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☐	L3	L2 same (attenuat$ or inactivat$ or weak$ or avirul$ or modifi$ or reduc$)	11
☐	L4	L2 same (attenuat$ or inactivat$ or weak$ or avirul$ or modifi$ or reduc$).ti,ab,clm.	11
☐	L5	l4 and temperature	3

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		DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI,TDBD; PLUR=YES; OP=AND	
☐	L1	brake.in. and neospor$	27
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☐	L3	L2 same (attenuat$ or inactivat$ or weak$ or avirul$ or modifi$ or reduc$)	11
☐	L4	L2 same (attenuat$ or inactivat$ or weak$ or avirul$ or modifi$ or reduc$).ti,ab,clm.	11
☐	L5	l4 and temperature	3
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END OF SEARCH HISTORY

L1: Entry 8 of 27 File: USPT Dec 2, 2003

US-PAT-NO: 6656479
DOCUMENT-IDENTIFIER: US 6656479 B2

TITLE: Attenuated live neospora vaccine

DATE-ISSUED: December 2, 2003

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Brake; David A	East Lyme	CT		
Blagburn; Byron L	Auburn	AL		
Lindsay; David S	Christiansburg	VA		

ASSIGNEE-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY	TYPE CODE
Pfizer Inc.	New York	NY			02

APPL-NO: 09/ 952388 [PALM]
DATE FILED: September 12, 2001

PARENT-CASE:
This application is a continuation of application Ser. No. 09/260,414, filed Feb. 26, 1999, now abandoned, which is a continuation of 08/967,744, filed Nov. 10, 1997, now abandoned, which claims priority from provisional application Ser. No. 60/031,248, filed Nov. 12, 1996 now abandoned.

INT-CL: [07] A61 K 39/002

US-CL-ISSUED: 424/269.1; 424/271.1, 424/273.1, 424/93.2, 424/93.1, 424/258.1, 435/69.1, 435/258.1, 800/947
US-CL-CURRENT: 424/269.1; 424/258.1, 424/271.1, 424/273.1, 424/93.1, 424/93.2, 435/258.1, 435/69.1

FIELD-OF-SEARCH: 424/93.1, 424/258.1, 424/271.1, 424/273.1, 424/93.2, 435/6, 435/69.1, 435/258.1, 800/947

PRIOR-ART-DISCLOSED:

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☐	5707617	January 1998	Conrad et al.	424/93.1

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☐	6071737	June 2000	Marsh et al.	

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FOREIGN-PAT-NO	PUBN-DATE	COUNTRY	US-CL
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ART-UNIT: 1645

PRIMARY-EXAMINER: Smith; Lynette R. F.

ASSISTANT-EXAMINER: Portner; Ginny Allen

ATTY-AGENT-FIRM: Scully, Scott, Murphy & Presser

ABSTRACT:

The present invention provides attenuated live cultures of the pathogenic protozoan parasite, Neospora, and live vaccines against neosporosis prepared therefrom which are useful in the prevention of clinical disease and abortion in mammals.

8 Claims, 0 Drawing figures

L1: Entry 8 of 27 File: USPT Dec 2, 2003

US-PAT-NO: 6656479
DOCUMENT-IDENTIFIER: US 6656479 B2

TITLE: Attenuated live neospora vaccine

DATE-ISSUED: December 2, 2003

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Brake; David A	East Lyme	CT		
Blagburn; Byron L	Auburn	AL		
Lindsay; David S	Christiansburg	VA		

US-CL-CURRENT: 424/269.1; 424/258.1, 424/271.1, 424/273.1, 424/93.1, 424/93.2, 435/258.1, 435/69.1

CLAIMS:

We claim:

1. A live culture of cells of an attenuated strain of a species of Neosyora, wherein the strain is NCTS-8 which is present in MARC145 monkey kidney cells having ATCC accession No. CRL12230.

2. A vaccine for protecting a mammal against neosporosis, comprising an immunologically effective amount of live cells of an attenuated strain of a species of Neosyora and a veterinarily acceptable carrier, wherein the strain is NCTS-8 which is present in MARC145 monkey kidney cells having ATCC accession No. CRL12230.

3. The vaccine of claim 2, further comprising an adjuvant.

4. The vaccine of claim 3, wherein the adjuvant is an oil-in-water emulsion.

5. A combination vaccine, comprising an immunologically effective amount of live cells of a an attenuated strain of a species of Neospora and a veterinarily acceptable carrier, wherein the strain is NCTS-8 which is present in MARC145 monkey kidney cells having ATCC accession No. CRL12230.

6. A composition comprising a veterinarily acceptable carrier and an immunologically effective amount of live cells of an attenuated strain of a species of Neosyora, wherein the strain is NCTS-8 which is present in MARC145 monkey kidney cells having ATCC accession No. CRL12230.

7. The composition of claim 6, further comprising an adjuvant.

8. The composition of claim 7, wherein the adjuvant is an oil-in-water emulsion.

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☐ 1. Document ID: US 20040081666 A1

L6: Entry 1 of 6 File: PGPB Apr 29, 2004

DOCUMENT-IDENTIFIER: US 20040081666 A1
TITLE: Cattle reproductive disease vaccines

Summary of Invention Paragraph:
[0009] Modified-live virus (MLV) vaccines, on the other hand, offer a higher level of protection. Currently, licensed BVDV MLV vaccines are produced using attenuated viruses obtained via repeated passage in bovine or porcine cells (Coggins et al., Cornell Vet. 51: 539-, 1961; Phillips et al., Am. J. Vet. Res. 36: 135-, 1975), or using chemically modifiedviruses which exhibit a temperature-sensitive phenotype (Lobmann et al., Am. J. Vet. Res. 45: 2498-, 1984; 47: 557-561, 1986). A single dose of MLV vaccine is sufficient for immunization, and duration of the immunity can last for years in vaccinated cattle. However, as these vaccines have been developed using type I BVDV virus strains, the protection is against type I virus only. Moreover, the available BVDV vaccines are not indicated for use in pregnant cattle or calves nursing pregnant cows.

Summary of Invention Paragraph:
[0022] The term "combination vaccine" is meant a bivalent or multivalent combination of antigens including modified live antigens and/or inactivated antigens. In accordance with the present invention a combination vaccine can comprise modified live infectious IBR, PI3, BRSV and inactivated BVDV Types 1 and 2, one or more antigens such as but not limited to Leptospira canicola, Leptospira grippotyphosa, Leptospira borgpetersenii hardio-prajitno, Leptospira icterohaemmorrhagia, Leptospira interrogans pomona, Leptospira borgpetersenii hardjo-bovis, Leptospira bratislava, Campylobacter fetus, Neospora caninum, Trichomonus fetus, Mycoplasma bovis, Haemophilus somnus, Mannheimia haemolytica and Pasturella multocida, a veterinary acceptable carrier and an adjuvant. In a preferred embodiment the modified live IBR component is temperature sensitive IBR. In another preferred embodiment the BVDV Type 2 component is cytopathic (cpBVD-2 strain 53637-ATCC No. PTA-4859) and the BVDV Type 1 component is cytopathic 5960 (cpBDV-1 strain 5960-National Animal Disease Center, United States Department of Agriculture, Ames, Iowa). The present invention also contemplates non-cytopathic BVDV Type 1 and Type 2 strains. In still another preferred embodiment, the modified live antigens are desiccated, lyophilized or vitrified.

CLAIMS:

18. A method of inducing an immune response against an antigen selected from the group consisting of Leptospira canicola, Leptospira grippotyphosa, Leptospira borgpetersenii hardio-prajitno, Leptospira icterohaemmorrhagia, Leptospira interrogans pomona, Leptospira borgpetersenii hardjo-bovis, Leptospira bratislava, Neospora caninum, Trichomonus fetus, Mycoplasma, bovis, Haemophilus somnus, Mannheimia haemolytica and Pasturella multocida in an animal subject, comprising

administering an immunologically effective amount of the composition of claim 1 and a veterinary-acceptable carrier.

54. A method of treating or preventing a disease or disorder in an animal caused by infection with an antigen selected from the group consisting Leptospira canicola, Leptospira grippotyphosa, Leptospira borgpetersenii hardio-prajitno, Leptospira icterohaemmorrhagia, Leptospira interrogans pomona, Leptospira borgpetersenii hardjo-bovis, Leptospira Bratislava, Campylobacter fetus, Neospora caninum, Trichomonus fetus, Mycoplasma, bovis, Haemophilus somnus, Mannheimia haemolytica and Pasturella multocida, comprising administering to said animal a therapeutically effective amount of the vaccine composition of claim 20.

2. Document ID: US 20030185852 A1

L6: Entry 2 of 6 File: PGPB Oct 2, 2003

DOCUMENT-IDENTIFIER: US 20030185852 A1
TITLE: Parasitic protozoan isolate

Abstract Paragraph:
The present invention relates to a novel Neospora caninum isolate from Nowra and extracts thereof. The strain is useful in the development of diagnostic assays for the detection of parasites in animals. The present invention also relates to pharmaceutical compositions, using live or killed organisms or extracts thereof, for the treatment and prevention of parasitic infections in animals.

Summary of Invention Paragraph:
[0012] The literature on live vaccines against N. caninum is limited. Atkinson et al. (1999) showed that infection of naive mice by the Nc-SweB1 isolate of N. caninum partially protected them against a severe infection by Nc-Liverpool. Lindsay et al. (1999) generated temperature sensitive mutants of N. caninum and demonstrated that they could prevent clinical signs associated with neosporosis in mice.

Detail Description Paragraph:
[0119] Lindsay D S, Lenz S D, Blagburn B L & Brake D A (1999) Characterization of temperature-sensitive strains of Neospora caninum in mice Journal of Parasitology 85, 64-67.

CLAIMS:

14. A method as claimed in any one of claims 10 to 13 wherein the infection or disease is caused by the presence of Neospora in the animal.

☐ 3. Document ID: US 20030091591 A1

L6: Entry 3 of 6 File: PGPB May 15, 2003

DOCUMENT-IDENTIFIER: US 20030091591 A1
TITLE: Alphavirus expression vectors and uses thereof

Detail Description Paragraph:
[0066] Sindbis virus gene expression, which occurs in the cytoplasm of the cell, is quite efficient, rapid, and can be modulated. For example, Xiong et al., ibid., reported the production of up to 1.times.10.sup.8 molecules of chloramphenicol acetyltransferase (CAT) per cell transfected with Sindbis virus expression vectors operatively linked to the CAT gene, when the cell was cultured for about 20 hr. Xiong et al. also reported that use of a replication temperature sensitive Sindbis virus vector led to modulated expression of CAT.

CLAIMS:

5. The vaccine of claim 1, wherein said disease is caused by an infectious agent selected from the group consisting of the genera Toxoplasma, Dirofilaria, Acanthocheilonema, Babesia, Brugia, Candida, Cryptococcus, Cryptosporidium, Dipetalonema, Eimeria, Encephalitozoon, Hepatozoon, Histoplasma, Isospora, Loa, Microsporidia, Neospora, Nosema, Onchocerca, Parafilaria, Plasmodium, Pneumocystis, Rochalimaea, Setaria, Stephanofilaria, Theileria and Wuchereria.

42. The recombinant molecule of claim 40, wherein said parasite is selected from the group consisting of Toxoplasma, Dirofilaria, Acanthocheilonema, Babesia, Brugia, Candida, Cryptococcus, Cryptosporidium, Dipetalonema, Eimeria, Encephalitozoon, Hepatozoon, Histoplasma, Isospora, Loa, Microsporidia, Neospora, Nosema, Onchocerca, Parafilaria, Plasmodium, Pneumocystis, Rochalimaea, Setaria, Stephanofilaria, Theileria and Wuchereria.

☐ 4. Document ID: US 20020102273 A1

L6: Entry 4 of 6 File: PGPB Aug 1, 2002

DOCUMENT-IDENTIFIER: US 20020102273 A1
TITLE: USE OF ALPHAVIRUS EXPRESSION VECTORS TO PRODUCE PARASITE ANITGENS

Detail Description Paragraph:
[0067] Sindbis virus gene expression, which occurs in the cytoplasm of the cell, is quite efficient, rapid, and can be modulated. For example, Xiong et al., ibid., reported the production of up to 1.times.10.sup.8 molecules of chloramphenicol acetyltransferase (CAT) per cell transfected with Sindbis virus expression vectors operatively linked to the CAT gene, when the cell was cultured for about 20 hr. Xiong et al. also reported that use of a replication temperature sensitive Sindbis virus vector led to modulated expression of CAT.

CLAIMS:

5. The vaccine of claim 1, wherein said disease is caused by an infectious agent selected from the group consisting of the genera Toxoplasma, Dirofilaria, Acanthocheilonema, Babesia, Brugia, Candida, Cryptococcus, Cryptosporidium, Dipetalonema, Eimeria, Encephalitozoon, Hepatozoon, Histoplasma, Isospora, Loa, Microsporidia, Neospora, Nosema, Onchocerca, Parafilaria, Plasmodium, Pneumocystis, Rochalimaea, Setaria, Stephanofilaria, Theileria and Wuchereria.

42. The recombinant molecule of claim 40, wherein said parasite is selected from the group consisting of Toxoplasma, Dirofilaria, Acanthocheilonema, Babesia, Brugia, Candida, Cryptococcus, Cryptosporidium, Dipetalonema, Eimeria, Encephalitozoon, Hepatozoon, Histoplasma, Isospora, Loa, Microsporidia, Neospora, Nosema, Onchocerca, Parafilaria, Plasmodium, Pneumocystis, Rochalimaea, Setaria, Stephanofilaria, Theileria and Wuchereria.

5. Document ID: US 6485904 B1

L6: Entry 5 of 6 File: USPT Nov 26, 2002

DOCUMENT-IDENTIFIER: US 6485904 B1
TITLE: DNA encoding a plasminogen activating protein

Detailed Description Text (132):
Following amplification with either ER43 (SEQ ID NO:22) plus ER45 (SEQ ID NO:24), or ER44 (SEQ ID NO:23) plus ER45 (SEQ ID NO:24), the triplicate samples were separately pooled by combining 7 .mu.l of each reaction mixture to produce a final volume of 21 .mu.l. The synthetic DNA fragments were separated in 1% (w/v) agarose and the 1,001 bp (ER44/ER45) and 1,073 bp (ER43/ER45) products were purified (JetSorb.TM., GenoMed, Research Triangle Park, N.C.). Following digestion of the purified DNA with KpnI and XbaI, approximately 0.3 .mu.g of the fragments were ligated to 0.5 .mu.g of KpnI plus XbaI restricted, dephosphorylated pEA181 vector DNA. Recombinant plasmids were transformed into competent E. coli TAP56 cells (Pfizer in-house strain) by the method of Hanahan (1983, J. Mol. Biol. 166: 557). Cells were grown in 1 ml SOC medium (Gibco BRL) for 75 min at 30.degree. C. Transformants were grown and maintained at .ltoreq.30.degree. C. to prevent induction of the P.sub.L promoter resulting from inactivation of the temperature-sensitive cI857 repressor. Two strains isolated in this manner were retained for expression of recombinant PauA. Strain Pz330 harbors plasmid pER330, which expresses the full-length pauA gene resulting from amplification of chromosomal DNA with primers ER43/ER45. Strain Pz332 harbors plasmid pER332, which expresses the truncated (signal peptide-deleted) pauA gene resulting from amplification with primers ER44/ER45.

CLAIMS:

42. The combination vaccine of claim 41, wherein the host cell is selected from the group consisting of Leptospira spp., Campylobacter spp., Staphylococcus spp., Streptococcus spp., Mycoplasma spp., Klebsiella spp., Salmonella spp., Pasteurella spp., Clostridium spp., E. coli, and Neospora spp.

43. The combination vaccine of claim 40, wherein the antigen of the second component of the combination vaccine is capable of inducing a protective response in the mammal against a disease, condition, or pathogen selected from the group consisting of mastitis, bovine herpes virus, bovine respiratory syncitial virus, bovine viral diarrhea virus, parainfluenza virus type I, parainfluenza virus type II, parainfluenza virus type III, Leptospira spp., Campylobacter spp., Staphylococcus spp., Streptococcus spp., Mycoplasma spp., Klebsiella spp., Salmonella spp., rotavirus, coronavirus, rabies, Pasteurella spp., Clostridium spp., Tetanus toxoid, E. coli, and Neospora spp.

46. The combination vaccine of claim 45, wherein the host cell is selected from the group consisting of Leptospira spp., Campylobacter spp., Staphylococcus spp., Streptococcus spp., Mycoplasma spp., Klebsiella spp., Salmonella spp., Pasteurella spp., Clostridium spp., E. coli, and Neospora spp.

47. The combination vaccine of claim 44, wherein the antigen of the second component of the combination vaccine is capable of inducing a protective response in the mammal against a disease, condition, or pathogen selected from the group consisting of mastitis, bovine herpes virus, bovine respiratory syncitial virus, bovine viral diarrhea virus, parainfluenza virus type I, parainfluenza virus type II, parainfluenza virus type III, Leptospira spp., Campylobacter spp., Staphylococcus spp., Streptococcus spp., Mycoplasma spp., Klebsiella spp., Salmonella spp., rotavirus, coronavirus, rabies, Pasteurella spp., Clostridium spp., Tetanus toxoid, E. coli, and Neospora spp.

☐ 6. Document ID: US 5766602 A

L6: Entry 6 of 6 File: USPT Jun 16, 1998

DOCUMENT-IDENTIFIER: US 5766602 A
TITLE: Recombinant packaging defective Sindbis virus vaccines

Detailed Description Text (25):
Sindbis virus vectors are preferred because, unlike Semliki Forest virus and other alphaviruses, Sindbis virus is not associated with human disease. In addition, Sindbis virus has a wide host range. For example, Sindbis virus can infect a number of organisms including mammalian, avian, insect, amphibian, and reptilian cells. Sindbis virus has infected all cell types studied so far, including, but not limited to, Chinese hamster ovary cells, baby hamster kidney cells, quail (e.g., QT-6) cells, chicken embryo fibroblasts, human tumor cells, mosquito and Drosophila cells. Sindbis virus can also be transmitted to vertebrate hosts, such as birds or mammals, by mosquitos. Sindbis virus gene expression, which occurs in the cytoplasm of the cell, is quite efficient, rapid, and can be modulated. For example, Xiong et al., ibid., reported the production of up to 1.times.10.sup.8 molecules of chloramphenicol acetyltransferase (CAT) per cell transfected with Sindbis virus expression vectors operatively linked to CAT gene in less than about 20 hr. The authors also reported that use of a replication temperature sensitive Sindbis virus vector led to modulated expression of CAT.

Other Reference Publication (40):
Rice et al., "Production of Infectious RNA Transcripts from Sindbis Virus cDNA

Clones: Mapping of Lethal Mutations, Rescue of a Temperature-Sensitive Marker, and in vitro Mutagenesis to Generate Defined Mutants", pp. 3809-3819, 1987, J. Virol., vol. 61, No. 12 (Dec.).

CLAIMS:

5. The method of claim 1, wherein said disease is caused by an infectious agent selected from the group consisting of the genera Toxoplasma, Dirofilaria, Cryptosporidium, Eimeria, Neospora, Isospora, Plasmoanum, Babesia, Theileria, Hepatozoon, Encephalitozoon, Nosema, Pneumocystis, Cryptococcus, Candida, and Histoplasma.

20. The method of claim 17, wherein said disease is caused by an infectious agent selected from the group consisting of the genera Toxoplasma, Dirofilaria, Cryptosporidium, Eimeria, Neospora, Isospora, Plasmodium, Babesia, Theileria, Hepatozoon, Encephalitozoon, Nosema, Pneumocystis, Cryptococcus, Candida, and Histoplasma.

Terms	Documents
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